

There are no translations available.

## **Of Mice and Men, and Chandeliers**

Alan Woodruff, Rafael Yuste

Citation: Woodruff A, Yuste R (2008) Of Mice and Men, and Chandeliers. PLoS Biol 6(9): e243. doi:10.1371/journal.pbio.0060243

Published: September 23, 2008

Copyright: © 2008 Woodruff and Yuste. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: This work was supported by the US National Eye Institute and the Kavli Institute for Brain Science.

Abbreviations: AAC, axo-axonic cell; EPSP, excitatory postsynaptic potential; IPSP, inhibitory postsynaptic potential

Alan Woodruff and Rafael Yuste are at the Howard Hughes Medical Institute, Department of Biological Sciences, Columbia University, New York, New York, United States of America.

### Of Mice and Men

What makes us human? One of the most obvious answers to this age-old question lies in the structure and function of the central nervous system, particularly the neocortex, where unique human features may lie. In fact, humans have not only a proportionally much larger neocortex compared to that of other mammals, but also a huge frontal lobe, the font of higher cognition.

In seeking clues to the biological basis of being human, neuroanatomists have long compared the human brain to that of other species, leading them to develop two distinct theories. Santiago Ramón y Cajal, the father of neuroanatomy, argued that the cortex of “higher” mammals, like humans, has more classes of neurons than those of “lower” mammals, for which he used the mouse as an example (1). Specifically, he proposed that the variety and sophistication of “short-axon” cells, i.e., GABAergic inhibitory interneurons, increases as one climbs up the evolutionary ladder (2).

The alternative position—that differences among species arise not from variations in cell types, but from the size and complexity of the circuits—was defended by Cajal's own disciple, Rafael Lorente de Nó, who, like many of the best students, did exactly the opposite of what he had been taught. Choosing the mouse as his experimental system for his thesis at the tender age of 20, Lorente described as many cell types in the mouse neocortex as Cajal had described in humans. Cajal politely published Lorente's paper in his journal without corrections (3), yet told his disciple that he was wrong. The argument would continue until Cajal's death: on his deathbed in 1934, Cajal wrote to Lorente, admonishing him: “the mouse is not a good choice for the study of cortical circuits because of its paucity of short-axon cells” (4).

### Of Chandelier Cells

One of the most distinct types of short-axon cells, or GABAergic interneurons, present in mammalian cortical circuits is the “chandelier” cell. Their distinct axonal arbor, with parallel arrays of short vertical sets of presynaptic terminals (“cartridges”), resembles the candlesticks of an old-fashioned chandelier. Chandelier neurons are rare, forming only a small percentage of all GABAergic interneurons (5)—both Cajal and Lorente missed them—and were not described

until 1974 by Szentagothai and Arbib (6). A similar neuron with parallel arrays of terminals (type 4 cell) was reported by Jones at about the same time (7). Based on the morphology of their terminals, Szentagothai thought that chandelier cells formed arrays of synapses with apical dendrites of pyramidal neurons (8), but this idea was proven wrong. Szentagothai's own disciple, Peter Somogyi, used electron microscopy to demonstrate that morphologically similar neurons, which he named axo-axonic cells (AACs), specifically contact the axon initial segment of pyramidal cells (9). This key finding was confirmed by Fairen and Valverde (10) and DeFelipe et al. (11), who proposed that chandelier cells and AACs were the same cell type. Both terms have been used interchangeably in the literature ever since.

The striking morphologies of chandelier neurons have captured the imagination of cortical researchers and are often used as the best examples to illustrate the apparently purposeful design of cortical microcircuits. Each chandelier cartridge establishes a large number of synapses with each pyramidal neuron, strategically placed in the axon initial segment, where the action potential is generated. Thus, chandeliers appear ideally suited to shut off entire groups of pyramidal cells, making them the ultimate cortical switches.

Until recently, little was known about the function of chandelier cells, owing both to their rarity and the lack of unique neurochemical or physiological markers. Occasional recordings from chandelier cells in vitro (12,13) and in vivo (14,15) revealed their interneuronal firing properties. But two years ago, a landmark paper by Gabor Tamás and colleagues (16) turned the field upside down. Tamás, himself a disciple of Somogyi, argued that chandelier cells have an excitatory as well as an inhibitory function. Amazingly, a single action potential in a chandelier neuron could directly drive multiple postsynaptic pyramidal cells to spike, providing a high-fidelity mechanism for signal propagation in a local cortical microcircuit. Forcing pyramidal cells to spike could result in excitatory feedback on the chandelier cells, providing a physiological marker to distinguish at least some chandelier cells from other interneurons. A similar phenomenon of feedback excitation had likely been seen in hippocampal chandelier cells over a decade earlier (12), and was also recently described in the amygdala (17), suggesting that the excitatory role of chandelier cells may in fact be widespread.

To explain how chandelier cells could be excitatory, Tamás and colleagues argued that the GABA reversal potential (EGABA) is more depolarized at the axon than elsewhere in the neuron, due to the lack of the potassium chloride cotransporter KCC2, which extrudes chloride to the extracellular space. But simply lacking the cotransporter may be insufficient to maintain such a large EGABA gradient between the axon and soma. Subsequently, a recent study utilizing GABA uncaging has reported a depolarizing shift in EGABA from dendrite to soma to the axon, and further showed that high axonal expression of the chloride importer NKCC1, perhaps in addition to a lack of KCC2, could maintain a depolarized EGABA (18).

### Of Human Chandeliers

In this issue of PLoS Biology, Molnár et al. extend their earlier findings on cortical chandelier cells, performing a technical tour de force. Recording from human surgical samples, the authors identify—for the first time in humans—pairs of connected neurons and study their synaptic and circuit properties (19). Dual recordings from connected cells are the current “gold standard” of circuit neuroscience, because they allow physiological analysis of the effect of activating single axons (20,21), and thus the functional isolation of elementary, neuron-to-neuron, synaptic responses.

Molnár et al. show that a single action potential in a single layer 2/3 cortical pyramidal cell can trigger polysynaptic chains of activity, detected as excitatory postsynaptic potentials (EPSPs) and inhibitory postsynaptic potentials (IPSPs) in recorded neurons. This reveals an extremely efficacious means of activity propagation in the cortical network. Although earlier work had shown polysynaptic activations following a single chandelier spike (16,17), the current study demonstrates much longer responses. Moreover, one of the most interesting results from Molnár et al. relates to the temporal structure of the activity patterns elicited after stimulation of a single neuron. While most of them appear to propagate through the circuit with increasing disorganization, occasionally the authors were able to trigger an amazingly precise temporal pattern. This implies that the microcircuit is capable under some circumstances of generating patterns of activation with low jitter and high temporal precision, resembling precise spatiotemporal patterns of network activation reported in neocortical preparations in vivo and in vitro (22-24).

What is the mechanism of these activations? The authors identify two key factors. First, very strong (up to 8–20 mV) connections from pyramidal cells to basket and chandelier cells enable spike-to-spike transmission. In human brain slices, a relatively high proportion of basket (20%) and chandelier neurons (33%) could be driven to threshold by a single pyramidal neuron spike—in stark contrast to an estimated 1% likelihood of finding polysynaptic events in rat. The second factor involves chandelier cell recruitment of downstream pyramidal cells through their depolarizing effect. These downstream pyramidal cells may thus produce EPSPs three synapses removed from the original spike, while—by virtue of large-amplitude synapses—perhaps activating more chandeliers and basket cells (thereby producing a second round of IPSPs, this time four synapses downstream of the original spike!). Such polysynaptic chains of activation, alternating between pyramidal and axo-axonic cells, could theoretically continue unabated, except that synapses onto, and from, many interneurons exhibit synaptic depression (a decrease in synaptic strength (25,26)).

The finding of such strong synapses between pyramidal cells and interneurons in human samples raises many questions. While the authors rule out interspecies differences in interneuron input resistance *in vitro*, it remains plausible that *in vivo* input resistances are lower for humans than for other animals. In this case, the strikingly large synaptic amplitudes may reflect an adaptation to different electrotonic conditions. Also, given that the activation of pyramidal neurons by chandelier cells depends crucially on the resting membrane potential of the pyramidal cell and the chloride equilibrium potential at the axon initial segment, it is possible that the exact extracellular milieu in which the neurons are bathed could facilitate, or impede, this effect. In this respect, it is essential to repeat these rodent experiments *in vivo*. Also, the preparation of human surgical samples differs significantly from that of rodent brain slices, and it is possible that the reported interspecies differences result from the different methods used. Alternatively, the large-amplitude EPSPs that enable extended polysynaptic sequences could indeed be unique to humans. Finally, even a relatively distinct group of neurons such as chandelier/axo-axonic cells is composed of cells with different morphologies (11), so it is conceivable that different subtypes of chandeliers or AACs exist. Indeed, a recent report has described a new type of AAC (27), raising the possibility that equating chandelier cells and AACs may be too simplistic. Therefore, one needs to re-examine whether all AACs are chandelier cells and to establish whether the findings of Molnár et al. (19) and Tamás et al. (16) apply to all chandeliers or AACs, or only to a subtype of them.

### Significance and Future Directions

Regardless of these potential issues, the data stand in front of us, providing us a tantalizing glimpse into the functional microcircuitry of the human neocortex, as well as a new chapter in the fascinating exposition of the chandelier cell. Moreover, the results of Molnár et al. cut to the heart of circuit neuroscience, addressing the basic question of how activity propagates through a circuit. Specifically, their data contribute to three important problems:

1. Role of single neurons in cortical networks: Given that even a small volume of neocortex contains tens of thousands of neurons, and that excitatory synapses are generally weak, with depressing dynamics and low probability of success, the role of individual neurons in the cortex (or more generally, in the mammalian brain) is thought to be negligible. Rather, it is traditionally believed that only the joint activity of many neurons can rise above these biophysical limitations to have any functional impact. But recent data from the Brecht laboratory have challenged this basic assumption (28,29). These *in vivo* experiments showed that the activation of individual neurons can alter the motor or sensory behavior of the animal, revealing the salience of single neuron in the brain. These remarkable experiments have lacked a mechanism that could explain how action potentials generated by a single neuron could ever propagate through these

biophysical hurdles. Molnár et al. now provide a potential mechanism for the Brecht data: perhaps the stimulated neurons trigger the activity of chandelier cells, lighting up chains of activity. At the same time, the Brecht data were obtained in rats, so it would appear inconsistent with the low probability of generating polysynaptic chains in rat brain slices discussed by Molnár et al. Alternatively, there may be a difference in propagation efficacy in vitro versus in vivo. Nevertheless, the Molnár et al. data confirm that, at least in some circumstances and in some species, stimulation of individual cortical neurons in vivo can generate an activity pattern that propagates through the circuit.

2. Generation of precise activity patterns: A second significant contribution of Molnár et al. relates to the debate of whether the cortex can generate spatiotemporal patterns of activity with great accuracy. As mentioned, spontaneously generated precise patterns of activity have been reported in vitro and in vivo (22,23,24), yet for every paper that reports such patterns, there appears to be at least two studies that negate their statistical significance. The fact that these precise patterns can actually be triggered by the experimenter, as shown by Molnár et al. (and also by MacLean et al. with thalamic stimulation (30)), makes the discussion of their statistical significance moot, and reorients the question to the examination of their mechanism and function. Leaving aside the potential function of these precise patterns for another discussion, the data of Molnár et al. could explain how these patterns are generated, by demonstrating that they can be triggered by the firing of single pyramidal neurons. This is a very different scenario from past proposals, which have focused on the synchronous firing of groups of cortical neurons (Abeles' synfire chains (31)), or on the pacemaker behavior found in subtypes of cortical cells (32). The demonstration that cortical circuits can generate and propagate precise spatiotemporal patterns of activity, together with the data from Brecht et al. eliciting stereotypical motor patterns by stimulating individual cortical neurons, supports the possibility that the cortex may fundamentally resemble the central pattern generators that dominate motor circuits (33,34), as if the forebrain represented the encephalization of more primitive fixed action patterns (35).

3. Human differences: Finally, the data from Molnár et al. reveal strong synaptic pathways in human neocortex. These strong interactions have not been seen before in other species, raising the possibility that human neocortex is endowed with specialized circuit properties. This is a controversial suggestion, since our large prefrontal lobes suggests that mental differences among species are due to differences in the size of cortical circuits, rather than differences in their neurons or modes of operation. Nevertheless, recordings from monkey interneurons have revealed physiological differences from those from rats, as if neurons from the same type were functionally different across species (36,37). In fact, although chandelier cells have been described in many mammalian species, including marsupials (38), they are particularly complex in humans, with larger axons and morphologically more elaborate cartridges, as compared to those of mice (5,39). One could argue that these morphological differences may translate into the ability to strongly recruit polysynaptic chains. Or, perhaps morphological differences between chandelier cells are coincidental, and it is the biophysical characteristics of human excitatory synapses onto interneurons that enable these uniquely human circuit properties. Finally, perhaps human cortical circuits have significant physical differences from those from other species, an idea supported by the morphological differences found when comparing the neocortex across different mammalian species (reviewed in (40)).

Further investigations of all these intriguing possibilities could potentially lead us to the essence of our humanity and mental world, or demonstrate, on the other hand, our similarities with other species. In any case, almost a hundred years after it started, the debate between Cajalians and Lorentians as to whether mice are essentially different than men is still open.

---

### References

1. Ramón y Cajal S (1899) La textura del sistema nerviosa del hombre y los vertebrados. 1st edition Madrid: Moya.
2. Ramón y Cajal S (1923) Recuerdos de mi vida: Historia de mi labor científica. Madrid: Alianza Editorial.
3. Lorente de Nó R (1922) La corteza cerebral del ratón. Trab Lab Invest Bio (Madrid) 20: 41–78.
4. Ramon y Cajal S (1934) Letter to Lorente. Courtesy of Dr Francisco Alvarez, translation by RY.
5. Inda MC, Defelipe J, Munoz A (2007) The distribution of chandelier cell axon terminals that express the GABA plasma membrane transporter GAT-1 in the human neocortex. Cereb Cortex 17: 2060–2071.
6. Szentagothai J, Arbib MA (1974) Conceptual models of neural organization. Neurosci Res Program Bull 12: 305–510.
7. Jones EG (1975) Varieties and distribution of non-pyramidal cells in the somatic sensory cortex of the squirrel monkey. J Comp Neurol 160: 205–267.
8. Szentagothai J (1975) The ‘module-concept’ in cerebral cortex architecture. Brain Res 95: 475–496.
9. Somogyi P (1977) A specific ‘axo-axonal’ interneuron in the visual cortex of the rat. Brain Res 136: 345–350.
10. Fairen A, Valverde F (1980) A specialized type of neuron in the visual cortex of cat: A Golgi and electron microscope study of chandelier cells. J Comp Neurol 194: 761–779.
11. DeFelipe J, Hendry SH, Jones EG, Schmechel D (1985) Variability in the terminations of GABAergic chandelier cell axons on initial segments of pyramidal cell axons in the monkey sensory-motor cortex. J Comp Neurol 231: 364–384.
12. Buhl EH, Han ZS, Lorinczi Z, Stezhka VV, Karnup SV, et al. (1994) Physiological properties of anatomically identified axo-axonic cells in the rat hippocampus. J Neurophysiol 71: 1289–1307.
13. Tamás G, Szabadics J (2004) Summation of unitary IPSPs elicited by identified

axo-axonic interneurons. *Cereb Cortex* 14: 823–826.

14. Klausberger T, Magill P, Marton L, Roberts J, Cobden P, et al. (2003) Brain-state- and cell-type-specific firing of hippocampal interneurons in vivo. *Nature* 421: 844–848.

15. Zhu Y, Stornetta RL, Zhu JJ (2004) Chandelier cells control excessive cortical excitation: Characteristics of whisker-evoked synaptic responses of layer 2/3 nonpyramidal and pyramidal neurons. *J Neurosci* 24: 5101–5108.

16. Szabadics J, Varga C, Molnar G, Olah S, Barzo P, et al. (2006) Excitatory effect of GABAergic axo-axonic cells in cortical microcircuits. *Science* 311: 233–235.

17. Woodruff AR, Monyer H, Sah P (2006) GABAergic excitation in the basolateral amygdala. *J Neurosci* 26: 11881–11887.

18. Khirug S, Yamada J, Afzalov R, Voipio J, Khiroug L, et al. (2008) GABAergic depolarization of the axon initial segment in cortical principal neurons is caused by the Na-K-2Cl cotransporter NKCC1. *J Neurosci* 28: 4635–4639.

19. Molnár G, Oláh S, Komlósi G, Füle M, Szabadics J, et al. (2008) Complex events initiated by individual spikes in the human cerebral cortex. *PLoS Biol* 6(9): e222.

20. Thomson AM, Girdlestones D, West DC (1988) Voltage-dependent currents prolong single-axon postsynaptic potentials in layer 3 pyramidal neurons in rat neocortical slices. *J Neurophysiol* 60: 1896–1907.

21. Markram H, Luebke J, Frotscher M, Sakmann B (1997) Regulation of synaptic efficacy by coincidence of postsynaptic APs and EPSPs. *Science* 275: 213–215.

22. Abeles M, Bergman H, Margalit E, Vaadia E (1993) Spatiotemporal firing patterns in the frontal cortex of behaving monkeys. *J Neurophysiol* 70: 1629–1638.

23. Mao BQ, Hamzei-Sichani F, Aronov D, Froemke RC, Yuste R (2001) Dynamics of spontaneous activity in neocortical slices. *Neuron* 32: 883–898.

24. Ikegaya Y, Aaron G, Cossart R, Aronov D, Lampl I, et al. (2004) Synfire chains and cortical songs: Temporal modules of cortical activity. *Science* 304: 559–564.

25. Xiang Z, Huguenard JR, Prince DA (2002) Synaptic inhibition of pyramidal cells evoked by different interneuronal subtypes in layer v of rat visual cortex. *J Neurophysiol* 88: 740–750.

26. Pouille F, Scanziani M (2004) Routing of spike series by dynamic circuits in the hippocampus. *Nature* 429: 717–723.

27. Gonchar Y, Turney S, Price JL, Burkhalter A (2002) Axo-axonic synapses formed by somatostatin-expressing GABAergic neurons in rat and monkey visual cortex. *J Comp Neurol* 443: 1–14.

28. Houweling AR, Brecht M (2008) Behavioural report of single neuron stimulation in somatosensory cortex. *Nature* 451: 65–68.

29. Brecht M, Schneider M, Sakmann B, Margrie TW (2004) Whisker movements evoked by stimulation of single pyramidal cells in rat motor cortex. *Nature* 427: 704–710.

30. MacLean JN, Watson BO, Aaron GB, Yuste R (2005) Internal dynamics determine the cortical response to thalamic stimulation. *Neuron* 48: 811–823.

31. Abeles M (1991) *Corticonics: Neural circuits of the cerebral cortex*. Cambridge: Cambridge University Press. 294 p.

32. LeBon-Jego M, Yuste R (2007) Persistently active, pacemaker-like neurons in primary visual cortex. *Front Neurosci* 1: 123–129.

33. Llinás RR (1988) The intrinsic electrophysiological properties of mammalian neurons: Insights into central nervous system function. *Science* 242: 1654–1664.



34. Yuste R, MacLean JN, Smith J, Lansner A (2005) The cortex as a central pattern generator. *Nat Rev Neurosci* 6: 477–483.
35. Llinás R (2002) *I of the vortex: From neurons to self*. Cambridge (MA): MIT Press.
36. Povysheva NV, Zaitsev AV, Kroner S, Krimer OA, Rotaru DC, et al. (2007) Electrophysiological differences between neurogliaform cells from monkey and rat prefrontal cortex. *J Neurophysiol* 97: 1030–1039.
37. Povysheva NV, Zaitsev AV, Rotaru DC, Gonzalez-Burgos G, Lewis DA, et al. (2008) Parvalbumin-positive basket interneurons in monkey and rat prefrontal cortex. *J Neurophysiol*. E-pub 16 July 2008.
38. Tyler CJ, Dunlop SA, Lund RD, Harman AM, Dann JF, et al. (1998) Anatomical comparison of the macaque and marsupial visual cortex: Common features that may reflect retention of essential cortical elements. *J Comp Neurol* 400: 449–468.
39. Inda MC, Defelipe J, Munoz A (2008) Morphology and distribution of chandelier cell axon terminals in the mouse cerebral cortex and claustroramygdaloid complex. *Cereb Cortex*. E-pub 24 April 2008.
40. (2007) Specializations of the cortical microstructure of humans. In: Kaas JH, Preuss TM, editors. *Evolution of the nervous systems: A comprehensive reference*. Amsterdam: Elsevier. pp. 167–190.